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Citation style: Sitek Patrycja, Chmielowska Aleksandra, Jaworska Maria, Lodowski Piotr, Szczepańska Marzena. (2019). Theoretical study of cobalt and nickel complexes involved in methyl transfer reactions: structures, redox potentials and methyl binding energies. "Structural Chemistry" Vol. 30, iss. 5, s. 1957-1970. DOI 10.1007/s11224-019-01384-z



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ORIGINAL RESEARCH



Theoretical study of cobalt and nickel complexes involved in methyl transfer reactions: structures, redox potentials and methyl binding energies

Patrycja Sitek¹ · Aleksandra Chmielowska¹ · Maria Jaworska¹ · Piotr Lodowski¹ · Marzena Szczepańska¹

Received: 25 March 2019 / Accepted: 25 June 2019 / Published online: 29 July 2019 © The Author(s) 2019

Abstract

Cobalamins, cobalt glyoximate complexes and nickel complexes with Triphos (bis(diphenylphosphinoethyl)phenylphosphine) and PPh₂CH₂CH₂SEt ligands were studied with the DFT/BP86 method in connection with methyl transfer reactions. Geometries, methyl binding energies and redox potentials were determined for the studied complexes. Three- and four-coordinate structures were considered for nickel complex with PPh₂CH₂CH₂SEt ligand, whereas four- and five-coordinate for its methyl derivative. On the basis of calculations, the possible mechanism of methyl transfer reaction between cobalt and nickel complexes was considered.

Keywords Nickel complexes · Cobalt complexes · DFT · Redox potentials · Methyl transfer

Introduction

The B_{12} vitamin derivatives (cobalamins) present in methyltransferases take part in many enzymatic methyl transfer reactions [1–3]. A unique methyl transfer reaction, where metals act as donors and acceptors of the methyl group, is found in the acetyl-CoA (Ljungdahl-Wood) pathway of autotrophic carbon fixation in various bacteria and archaea [4]. Acetyl-CoA is synthesized at the Ni-Ni-[4Fe-4S] cluster (the A-cluster) of acetyl-CoA synthase (ACS) through condensation of coenzyme-A (CoASH) with CO and the methyl group from CH₃-Cob(III)alamin of the corrinoid-iron-sulfur protein (CoFeSP) [5, 6]. A key step of such synthesis is the transfer of the methyl group from CoFeSP to the proximal Ni atom in the active site of ACS [7]. This reaction proceeds according to the equation:

$$CH_3 - Co(III)FeSP + CO + CoASH \rightleftharpoons CH_3CO$$

- SCoA + Co(I)FeSP + H⁺ (1)

Electronic supplementary material The online version of this article (https://doi.org/10.1007/s11224-019-01384-z) contains supplementary material, which is available to authorized users.

The occurrence of Ni(0) [8, 9] or Ni(I) [10] in reaction (1) of ACS was postulated. Since the mechanism of catalytic action of the ACS enzyme is not fully understood [8, 11–13], models of methylation reactions involving nickel complexes and various methylation factors are being examined experimentally [9, 14–19]. Likewise, many complexes relevant to ACS enzyme are investigated experimentally [20–24] and computationally [25–29]. Examples of methylation reactions with nickel participation are [10, 28]:

$$Ni(Triphos)PPh_{3}+CH_{3}-Co(dmgBF_{2})_{2}py+sol \longrightarrow$$
$$Ni(Triphos)CH_{3}^{+}+Co(dmgBF_{2})_{2}sol^{-}+PPh_{3}+py,$$
(2)

where Triphos stands for bis(diphenylphosphinoethyl)phenylphosphine ligand, and

$$Ni(PPh_2CH_2CH_2SEt)_2 + CH_3J \longrightarrow$$

$$Ni(PPh_2CH_2CH_2SEt)_2CH_3^+ + J^-.$$
(3)

In general, two mechanisms— $S_N 2$ and radical—are possible in methyl transfer reactions with cobalamin participation [28, 30, 31]. Both reactions (2) and (3) involve methylation of nickel(0) complexes. In the first step of the radical reaction, the methyl derivative should be reduced by the methyl acceptor; thus, the homolytic cleavage of Co-CH₃ bond is initiated by electron transfer between reactants. The radical mechanism is therefore possible when methyl acceptor is able to reduce methyl donor.

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For theoretical modelling of such reactions with the use of DFT method, it is extremely important to apply a functional which properly describes electronic structure of the reactants, giving the results comparable with experiment. This is especially important for methyl-metal binding energy and oxidation-reduction properties of the reacting complexes. In this work, the calculational study was carried out for nickel and cobalt complexes pertinent to biological (1) and model (2), (3) methyl transfer reactions with the use of DFT method and nonhybrid functional BP86. This functional allows to get a good description of the cobalt-methyl bond in alkylcobalamins, while the hybrid functionals significantly underestimate the energy of this bond [32–35]. Calculations for transition metal complexes show that BP86 functional gives also good estimation for redox potentials [36–38]. In the present study, BP86 is used to determine geometry, methyl binding energies and redox potentials for the investigated cobalt and nickel complexes. The results are compared with the experimental data.

Method of calculations

The calculations were carried out with the use of Gaussian16 program [39]. The DFT method with BP86 [40, 41] functional and TZVP [42] basis sets were used in the calculations. The effect of environment was taken into account by PCM solvent model [43–45], with water (for Co complexes) and acetonitrile (for Ni complexes) as solvents. The geometry of all studied complexes was fully optimized. The zero point energy (ZPE) and G3 dispersion correction [46, 47] were added to the calculated binding energies.

Methylcobalamin (MeCbl) is a methyl derivative of vitamin B_{12} which is uses in methyltransferases enzymes as a methylation factor. In CoFeSP protein it exists as a base-off form. The base-on form of methylcobalamin has dimethylbenzimidazole (DMB) as a ligand trans to methyl [2]. In the base-off form, DMB is replaced by a water ligand. The base-on and base-off forms are shown in Fig. S1 (Supporting Material).

In the present study, the base-on and base-off structures were examined in the form of simplified models, in which all the corrin (denoted further as Cor) side chains were replaced by hydrogen atoms and for base-on methylcobalamin the 5,6-dimethylbenzimidazole trans axial base was replaced by imidazole [48]. The base-off form without trans axial base and with water molecule as ligand was also considered. The calculations for cobalt complexes without methyl ligand were also performed for the sake of comparison with the experimental data.

The calculations for cobalt dimethylglyoxime complexes and nickel complexes were carried out, in reference to the reactions (2) and (3). The Ni(PPh₂CH₂CH₂SEt)₂ complex was examined with nickel in three oxidation states: Ni(II), Ni(I) and Ni(0) which were studied experimentally [10]. In the case of the one and two-electron reduced complex the four and three coordinated complexes were investigated. For Ni(Triphos)PPh₃ complex, the calculations were performed in the neutral and oxidized state. The relevant methyl derivatives of nickel complexes were also studied.

Results and discussion

The aim of this work is to compare the properties of nickel and cobalt methyl derivatives in relation to methyl transfer reactions. It is essential in context of reactions (1) and (2), where the methyl group is transferred between the two metal centers, from cobalt to nickel. The important question is what properties of these complexes cause the reaction to run in this direction, what is the relative strength of methyl binding and other electron properties. It could help to explain the occurrence of nickel in the ACS enzyme and the unique biological properties of ACS enzyme. To the best of our knowledge, there are no studies with theoretical methods for methyl nickel complexes in the literature. The cobalamin-methyl complexes were extensively investigated theoretically [3, 32, 33, 35, 49-61] due to their enormous significance in biological processes. We performed also the calculations for cobalt complexes to have consistent data set obtained with the same method, basis set, solvent and other computational conditions.

Structure

The axial ligand distances for cobalt complexes and methylnickel bond lengths for nickel complexes are collected in Table 1. The obtained structures are presented in Figs. 1, 2, 3, 4, and 5. In Supporting Information, the total energies and selected geometrical parameters of the investigated complexes are given in Table S1 and Tables S2, S3, S4, and S5, respectively.

Cobalt complexes

For cobalamin and dimethylglyoxime complexes the geometry of optimized structures are shown in Figs. 1 and 2, respectively. The axial ligand bond lengths are gathered in Table 1. Other selected geometrical parameters for cobalamins and cobaloximes are presented in Tables S1 and S2, respectively and the numbering of atoms in Fig. S2.

For cobalamins, the most notable features are related to geometry changes occurring upon reduction. After reduction of the five-coordinated complex CoCorIm, the imidazole ligand dissociates and the four-coordinated CoCor(I) complex is formed. This is in agreement with the experimental Table 1 Selected distances (Å) in Co and Ni complexes

			Calc.	Exp.
CH ₃ CoCorIm ⁺		Co-C _{CH3}	1.990	1.972 ^a
		Co-N _{Im}	2.178	2.093 ^a
CH ₃ CoCorIm		$Co-C_{CH_3}$	1.981	
		Co-N _{Im}	2.169	
CH ₃ CoCor ⁺		Co-C _{CH3}	1.971	
CH ₃ CoCor		Co-C _{CH3}	1.959	
CoCorIm ⁺		Co-N_{Im}	2.160	
CoCorIm		Co-N _{Im}	19.745	
CH ₃ CoCorH ₂ O ⁺		Co-C _{CH3}	1.974	
		Co-O_{H_2O}	2.370	
CH ₃ CoCorH ₂ O ^d		Co-C _{CH3}	1.960	
		Co-H2 _{<i>H</i>2<i>O</i>}	3.073	
		N1-H1 _{H2O}	2.588	
		N2-H2 _{H2O}	2.654	
CoCorH ₂ O ⁺		Co-O_{H_2O}	2.304	
CoCorH ₂ O		Co-H_{H_2O}	2.218	
CH ₃ Co(dmgBF ₂) ₂ py		Co-C _{CH3}	2.033	2.007 ^b
		Co-N_{py}	2.082	2.119 ^b
CH ₃ Co(dmgBF ₂) ₂ py ⁻		Co-C _{CH3}	2.003	
		Co-N_{py}	2.269	
CH ₃ Co(dmgBF ₂) ₂		Co-C _{CH3}	1.980	
$CH_3Co(dmgBF_2)_2^-$		Co-C _{CH3}	2.001	
Co(dmgBF ₂) ₂ py		Co-N_{py}	2.050	
$Co(dmgBF_2)_2 py^-$		Co-N_{py}	1.993	
$CH_3Ni(PPh_2CH_2CH_2SEt)_2^+$	MeVI ^e	Ni-C _{CH3}	1.981	
CH ₃ Ni(Triphos) ⁺		Ni-C _{CH3}	1.975	1.963 ^c

^aRef. [82] ^bRef. [19]

^cRef. [28]

^dNumbering of atoms is presented in Fig. S1

^eFor the lowest energy conformer (Fig. 4 and Table S1)

data and theoretical calculation results [3, 48, 51, 62–64]. In the reduced base-off methylcobalamin with a water molecule as an axial ligand the water is coordinated to cobalt by the oxygen atom. The reduction of $CH_3Co(III)CorH_2O$ leads to a system with water linked by hydrogen bond to corrin nitrogens ($CH_3Co(II)CorH_2O$). In contrast to that, in the reduced methyl-free cobalt complex $CoCorH_2O$, the water molecule is bound by hydrogen bond to the cobalt atom (see Fig. 1 and Table 1). The existence of cobalt-water hydrogen bonding was predicted theoretically [64].

In the dimethylglyoxime complexes, the axial base coordinated to cobalt is pyridine (Fig. 2). The BP86 results reveal that upon reduction pyridine is not detached both in methylated and methyl-free complexes.

Nickel complexes

The structures of nickel complexes are depicted in Figs. 3, 4, 5, whereas methyl-nickel bond lengths and other selected optimized geometrical parameters are collected in Tables 1, S4, and S5.

For Ni(PPh₂CH₂CH₂SEt)²⁺₂ a planar structure was obtained (Fig. 3, structure I^{2+}) which is in agreement with the crystal structure [10, 15]. The one- and two-electron reduced complexes I^+ and I are characterized by a distorted tetrahedral coordination (Fig. 3 and Table S3). For the two-electron reduced molecule Ni(PPh₂CH₂CH₂SEt)₂ which is a Ni(0) complex, the possibility of ligand dissociation was suggested [15]; hence, the calculations for three-coordinated



Fig. 1 The optimized structures of cobalamin complexes: ab, cd, kl reduction does not change the geometry of the complexes; ef – after reduction the imidazole ligand is detached; gh – after reduction the

forms of the one- and two-electron reduced complexes were also performed (II^+ - III^+ and II-III). II and III differ in the nickel coordination mode, in II nickel is coordinated by two phosphorus and one sulfur atom while in III by two sulfur and one phosphorus atom (Fig. 3 and Fig. S3). Optimized geometry reveals that the coordination of the nickel atom in two-electron reduced three-coordinate structures approximately corresponds to vertices of an almost equilateral triangle, while in the case of One-electron reduced twocoordinate phosphorus or sulfur atoms are almost linear (Fig. 3 and Table S3).

The computed energies (Fig. 3) show that the lowest energy complex is a four-coordinate one for both reduced states, I^+ and I. The three-coordinated complex with two phosphines (II) is 6.9 kcal/mol higher in energy (14.3 kcal with dispersion correction) than four-coordinate I. The three-coordinate complexes in which the nickel atom is coordinated by two sulfur atoms and one phosphorus are much higher in energy.

The methylated complex $CH_3Ni(PPh_2CH_2CH_2SEt)_2^+$ was examined in the form of five- and four-coordination structures. The obtained structures **MeI–MeVI** are shown in Fig. 4. These structures differ in mutual position of sulfur and phosphorus atoms and metal coordination number where **MeI** and **MeII** are five-coordinate and **MeII–MeVI** are four-coordinate. The lowest energy structure is

water ligand is bonded by hydrogen bond with nitrogen atoms of the corrin ring; \mathbf{ij} – after reduction the water ligand is bonded by hydrogen bond with the cobalt atom

MeVI, where nickel is coordinated by two phosphorus and one sulfur atoms and where sulfur is in trans position to the methyl group. BP86 functional gives five-coordinated structure, **MeII**, as a second one in energy (6.2 kcal/mol, and 1.6 kcal/mol higher with dispersion correction). Basing on the NMR spectra, the fivecoordinated geometry is suggested [10]. Taking into account a small energy difference calculated with D3 correction, it is possible that sulfur ligands undergo very fast exchange process.

The nickel complexes with Triphos ligand are depicted in Fig. 5 and the optimized geometry parameters are gathered in Table S5. The Ni(Triphos)PPh₃ which is a Ni(0) complex has a distorted tetrahedral structure, which is in agreement with the crystal structure [28]. Similarly the distorted tetrahedral geometry was obtained for Ni(Triphos)PPh₃⁺. The methyl derivative, CH₃Ni(Triphos)⁺ which is a Ni(II) complex, has a planar structure, which is also in accordance with the experiment [28].

Methyl-metal bonding

The methyl binding energy in the investigated cobalt and nickel complexes was computed according to the formula:

$$E_B = \Delta E + ZPE + D3 + BSSE, \tag{4}$$



Fig. 2 The optimized structures of dimethylglyoxime cobalt complexes

where

$$\Delta E = E(CH_3) + E(ML) - E(CH_3 - ML)$$

and ML, ZPE, D3 and BSSE denote metal complex, zero point energy, dispersion correction and basis set superposition error correction, respectively.

The obtained results are collected in Table 2 with the experimental data for comparison, where available.

There were many theoretical studies in which Co-C binding energy was calculated [3, 32–35, 49–61, 64–74]. It was shown that gradient functional BP86 gives binding energy close to experimental, while the hybrid functionals significantly underestimate its value. The TPSS functional was found to perform well in binding energy calculations for adenosylcobalamin system [73, 74]. We use BP86 as it gives good results for E_B and reduction potentials as well, as shown further. As mentioned earlier our binding energy calculations for cobalamins are performed to have consistent data set of computational results allowing for systematic comparison between cobalt and nickel complexes. Inspection of data in Table 2 reveals that the binding energies without dispersion correction are generally underestimated. The good agreement of the BP86+D3 calculated binding energies for methylcobalamin and its reduced form with experimental data [59, 75] is also found in the present calculations. The BSSE error is rather small (about 1.5 kcal) and of similar value for all complexes studied.

From the data in Table 2, it can be noted that for the reduced cobalamins (**3** and **5**, Table 2), the methyl binding energy is smaller than in the case of oxidized ones. The mechanism of methyl dissociation in the reduced methylcobalamin was studied theoretically, and it was shown that the reduction occurs on the corrin ring [76]. When looking at the spin density values collected in Table 3, one can find that indeed the unpaired electron is localized on the corrin ring in the reduced methylcobalamin. Similar pattern emerges from the electron density difference plots shown in Fig. 6, where the largest values are found on corrin carbons in the reduced methylcobalamin base-on and baseoff forms. As mentioned in the "Cobalt complexes" section, the reduced cobalamin occurs in the base-off form where the axial base is missing or replaced by a water molecule.

The results from the calculations show that for dimethylglyoxime cobalt complex with the axial pyridine ligand the Co-CH₃ bond energy is somewhat larger (about 3.5 kcal) than in methylcobalamin. After the reduction methyl binding energy decreases of about 10 kcal (to 33 kcal, Table 2). Unlike as in cobalamins, after reduction of Co(dmgBF₂)₂py, the pyridine ligand is not detached. When the pyridine ligand is missing (for 8 and 9), the methyl-cobalt binding energy for the oxidized and reduced forms are very similar (about 40 kcal), which is also different than in the case of MeCbl. These differences can be explained by inspecting the spin densities of the reduced cobalt complexes gathered in Table 3. The spin densities in reduced glyoximate complexes show that reduction occurs partially on the dimetyloglyoxime ligand and partially on cobalt atom, which is in contrast to cobalamins, where it occurs solely on corrin. This can be traced to more negatively charged cobaloxime than corrin ring (-2 vs. -1). This is also visible in Fig. 6 where pronounced values of electron density differences are present on cobalt and methyl. The calculated methyl binding energy in 6 from Table 2 (42 kcal) is larger than measured for CH₃Co(dmgh)₂py [52] amounting to 33.1 kcal; on the other hand, it is close to calculated value of 41.06 kcal for CH₃Co(dmgH)₂NH₃ [77] and 40.7 kcal for CH₃Co(dmgH)₂NHCH₂ [68].

For the nickel complex with the $PPh_2CH_2CH_2SEt$ ligand, the methyl binding energy is given only for the lowest energy conformer (**MeVI** in Fig. 3 and in Table S1),



Fig. 3 The optimized structures of nickel complexes with PPh₂CH₂CH₂SEt ligand. ΔE (in kcal/mol) denotes relative energy obtained from BP86 optimization, in parentheses the G3 correction is taken into account

and it amounts to 50.1 kcal/mol. The Ni-CH₃ bond energy calculated for the CH₃Ni(Triphos)⁺ complex is 52.3 kcal/mol and is the largest among all calculated E_B values. To the best of our knowledge, the nickel–methyl binding energies were not determined experimentally. For both nickel complexes, the calculated methyl binding energies are larger than those for cobalt cobaloximes and cobalamins. This accounts for the fact that the methyl is transferred from cobalt to nickel complex. In Table 4, the NBO bonding analysis for metal-methyl bond is given for cobalt and nickel complexes. Concerning the cobalt complexes it can be seen that the bonding σ_{Co-CH_3} orbital has approximately equal percentage participation of cobalt and carbon hybridized atomic orbitals (between 47% and 53%). The larger deviation is for CH₃Co(dmgBF₂)₂ with 43% and 57% cobalt and carbon orbital participation, respectively. For nickel complexes, the metal contribution to the bonding orbital amounts to 35%



Fig. 4 The optimized structures of methyl-nickel $CH_3Ni(PPh_2CH_2SEt)_2^+$ complex. ΔE (in kcal/mol) denotes relative energy obtained from BP86 optimization, in parentheses the G3 correction is taken into account

and carbon to 65%. Thus, some ionic character in Ni– CH₃ bonding can be inferred with participation of formally CH₃⁻ ion. In turn, the cobalt–methyl bond can be viewed as basically covalent.

In Table S5, NBO charges are collected for methyl-nickel and methyl-cobalt complexes. For cobalt complexes, the charge on the metal and the methyl group is positive, except for the reduced base-on glyoxime complex, where the metal is negative. The charge on metal and methyl in reduced and non-reduced cobalamin complexes is practically the same, indicating that the reduction occurs predominantly on the corrin ring. On the other hand, metal and methyl group are both more negative in the cobalt glyoximate reduced complexes. This corroborates with data in Table 3 and confirms the different behaviors of glyoximate and corrinate cobalt-methyl complexes upon reduction. These differences are due to the charge of the macrocyclic ligand, minus one for corrin and minus two for glyoxime, so the corrin ligand can accept a larger charge as a result of the reduction. In turn, in the methyl nickel complexes, the methyl group and nickel have negative charges.

Redox potentials

Redox potentials were calculated according to the equation:

$$E_{redox} = E(M^{n+1})_{sol} - E(M^n)_{sol} - 4.34(SHE)$$
(5)

The value of standard hydrogen electrode potential was taken from [78]. The obtained results are summarized in Table 5 and compared with experimental data. Because redox potentials were measured with the use of different reference electrodes, we converted all values to the standard hydrogen electrode (SHE). There were several measurements of redox potentials for various cobalamin forms [72, 75, 79–81]. The cobalamin redox potentials were also calculated theoretically [56, 64]. The calculations performed with BP86 functional show that it gives good





results for redox potentials of transition metal complexes [36–38].

Generally, it can be noted (Table 5) that the BP86 calculated redox potentials are in good agreement with the experimental data (maximum difference up to 0.2 eV). From the results it can be seen that for base-off cobalamins the redox potentials are more positive that for the base-on

ones. For cobaloxime complexes, calculated values of redox potentials are significantly more positive in comparison with similar forms of cobalamin complexes.

In regard to the reactions (2) and (3), there may be $S_N 2$ or radical mechanisms involved, the latter one with electron transfer from the nickel complex to methylcobalamin or methyl derivative of cobalt dimethylglyoximate. Looking

	Molecule	ΔE +ZPE	$\Delta E+ZPE$ +D3	$\Delta E+ZPE$ +D3+BSSE	Exp.
1	CH ₃ CoCorIm ⁺	30.4	40.0	38.7	$37\pm3^{a}, 36\pm4^{b}, 39\pm5^{c}$
2	CH ₃ CoCor ⁺	34.9	44.4	43.0	
3	CH ₃ CoCor	13.0	21.7	20.2	
4	CH ₃ CoCorH ₂ O ⁺	32.1	41.4	40.0	$44.6^{\rm d}, 42 \pm 5^{\rm c}$
5	CH ₃ CoCorH ₂ O	9.6	19.8	18.4	19.0 ^e
6	CH ₃ Co(dmgBF ₂) ₂ py	33.8	43.5	42.2	33.1 ^f
7	CH ₃ Co(dmgBF ₂) ₂ py ⁻	19.0	34.4	33.0	
8	CH ₃ Co(dmgBF ₂) ₂	32.5	41.1	39.7	
9	$CH_3Co(dmgBF_2)_2^-$	31.5	39.7	38.1	
10	CH ₃ Ni(PPh ₂ CH ₂ CH ₂ SEt) ^{+g} ₂	40.0	51.8	50.1	
11	CH ₃ Ni(Triphos) ⁺	44.5	54.6	52.9	

Table 2 Methyl binding energy E_B (kcal/mol)

^aRef. [60]

^bRef. [83] ^cRef. [84] ^aGas phase, Ref. [61] ^eRef. [75] ^fFor CH₃(dmgH)₂py, Ref. [52] ^gFor the lowest energy conformer **MeVI** (Fig. 4 and Table S1)

Co		0.049 0.044	0.004
$CH_3Co(dmgBF_2)_2py^0.300$ $CH_2Co(dmgBF_2)0.333$	$CH_3Co(dmgBF_2)_2py^-$ $CH_3Co(dmgBF_2)$	$\begin{array}{ccc} -0.048 & -0.944 \\ Co & (dmgBF_2)_2 \\ -0.300 & -0.644 \\ -0.333 & -0.590 \end{array}$	0.004 CH ₃ 0.023 -0.077

Fig. 6 Cross-sectional contours along the axial bonding for electron density difference between the oxidized and reduced form of selected cobalamin and dimethylglyoxime cobalt complexes, contour values between – 0.001 a.u (blue) and 0.001 a.u. (red)



Table 4 NBO analysis of axial bonds for cobalamin and dimethylglyoxime cobalt complexes (the hybridization of the atoms is indicated with the percent contribution of the metal-centered d or (and) p orbitals as a superscript, LP denotes an electron lone pair)

NBO	Occupancy	
CH ₃ CoCorIm ⁺		
$\sigma_{Co-C_{CH_3}}$	1.8093	$[47\%]0.6848(sp^{13.19}d^{54.29})_{Co} + [53\%]0.7287(sp^{81.11})_C$
$\sigma^*_{Co-C_{CH_2}}$	0.1326	$[53\%]0.7287(sp^{13.19}d^{54.29})_{Co} - [47\%]0.6848(sp^{81.11})_C$
$LP(N_{Im})$	1.6858	sp ^{62.99}
CH ₃ CoCorH ₂ O ⁺		
$\sigma_{Co-C_{CH_2}}$	1.8167	$[49\%]0.7033(sp^{8.29}d^{57.87})_{Co} + [51\%]0.7109(sp^{82.37})_{Co}$
$\sigma^*_{Co-C_{CH_2}}$	0.1161	$[51\%]0.7109(sp^{8.29}d^{57.87})_{Co} - [49\%]0.7033(sp^{82.37})_{Co}$
$LP(O_{H_2O})$	1.9908	<i>sp</i> ^{69.75}
$LP(O_{H_2O})$	1.8715	sp ^{76.39}
CH ₃ CoCor ⁺		
$\sigma_{Co-C_{CH_2}}$	1.8258	$[51\%]0.7176(sp^{4.43}d^{60.81})_{Co} + [49\%]0.6965(sp^{83.72})_{Co}$
$\sigma^*_{Co-C_{CH_3}}$	0.1245	$[49\%] 0.6965 (sp^{4.43}d^{60.81})_{Co} - [51\%] 0.7176 (sp^{83.72})_C$

Table 4(continued)

NBO	Occupancy	
CH ₃ Co(dmgBF ₂) ₂ py		
$\sigma_{Co-C_{CH_3}}$	1.6952	$[43\%]0.6594(sp^{45.18}d^{38.00})_{Co} + [57\%]0.7518(sp^{81.67})_C$
$\sigma_{Co-N_{py}}$	1.8863	$[17\%]0.4147(sp^{54.68}d^{29.42})_{Co} + [83\%]0.9099(sp^{71.82})_N$
$\sigma^*_{Co-C_{CH_3}}$	0.0805	$[57\%] 0.7518 (sp^{45.18}d^{38.00})_{Co} - [43\%] 0.6594 (sp^{81.67})_C$
$\sigma^*_{Co-N_{py}}$	0.1686	$[83\%] 0.9099 (sp^{54.68}d^{29.42})_{Co} - [17\%] 0.4147 (sp^{71.82})_N$
CH ₃ Co(dmgBF ₂) ₂		
$\sigma_{Co-C_{CH_3}}$	1.8241	$[52\%]0.7203(sp^{5.13}d^{61.09})_{Co} + [48\%]0.6936(sp^{84.94})_C$
$\sigma^*_{Co-C_{CH_3}}$	0.0691	$[48\%] 0.6936 (sp^{5.13}d^{61.09})_{Co} - [52\%] 0.7203 (sp^{84.94})_C$
$CH_3Ni(PPh_2CH_2CH_2SEt)_2^{+a}$		
$\sigma_{Ni-C_{CH_3}}$	1.7679	$[35\%]0.5907(sp^{42.99}d^{34.48})_{Ni} + [65\%]0.8069(sp^{79.18})_C$
$\sigma_{Ni-S_{trans}}$	1.9082	$[20\%]0.4481(sp^{56.68}d^{22.53})_{Ni} + [80\%]0.8940(sp^{80.19})_S$
$\sigma^*_{Ni-C_{CH_2}}$	0.1237	$[65\%] 0.8069 (sp^{42.99}d^{34.48})_{Ni} - [35\%] 0.5907 (sp^{79.18})_C$
$\sigma^*_{Ni-S_{trans}}$	0.1295	$[80\%] 0.8940 (sp^{56.68}d^{22.53})_{Ni} - [20\%] 0.4481 (sp^{80.19})_S$
CH ₃ Ni(Triphos) ⁺		
$\sigma_{Ni-C_{CH_3}}$	1.7856	$[35\%]0.5894(sp^{42.73}d^{33.16})_{Ni} + [65\%]0.8078(sp^{79.23})_C$
$\sigma_{Ni-P_{trans}}$	1.8443	$[28\%]0.5258(sp^{57.18}d^{22.48})_{Ni} + [72\%]0.8506(sp^{70.42})_P$
$\sigma^*_{Ni-C_{CH_2}}$	0.1290	$[65\%] 0.8078 (sp^{42.73}d^{33.16})_{Ni} - [35\%] 0.5894 (sp^{79.23})_C$
$\sigma^*_{Ni-P_{trans}}$	0.1311	$[72\%]0.8506(sp^{57.18}d^{22.48})_{Ni} - [28\%]0.5258(sp^{70.42})_P$

 aFor the lowest energy conformer (MeVI, Fig. 4 and Table S1)

Table 5Redox potentials E_{redox} (V)

		Calculated	Exp.	SHE	ΔE_0^{a}
CH ₃ CoCorIm ⁺ /CH ₃ CoCorIm		- 1.58	$-1.60^{b,f}$	- 1.36	(-0.22)
CH ₃ CoCor ⁺ /CH ₃ CoCor		-1.41	- 1.45 ^{b,g}	- 1.21	(-0.20)
CoCorIm ⁺ /CoCorIm		-0.78	$-0.85^{b,g}$	-0.61	(-0.17)
CH ₃ CoCorH ₂ O ⁺ /CH ₃ CoCorH ₂ O		-1.40			
CoCorH ₂ O ⁺ /CoCorH ₂ O		- 0.36	$-0.74^{b,g}$	-0.50	(0.14)
CoCor ⁺ /CoCor		-0.38			
CH ₃ Co(dmgBF ₂) ₂ py/CH ₃ Co(dmgBF ₂) ₂ py ⁻		-0.99	- 1.10 ^{c,h}	-1.10	(0.11)
$CH_3Co(dmgBF_2)_2/CH_3Co(dmgBF_2)_2^-$		-0.15			
Co(dmgBF ₂) ₂ py/Co(dmgBF ₂) ₂ py ⁻		-0.14			
$Co(dmgBF_2)_2/Co(dmgBF_2)_2^-$		-0.08	$-0.55^{b,j}$	- 0.31	(0.23)
Ni(PPh ₂ CH ₂ CH ₂ SEt) ²⁺ /Ni(PPh ₂ CH ₂ CH ₂ SEt) ⁺		0.01	$-0.56^{d,i}$	-0.02	(0.03)
Ni(PPh ₂ CH ₂ CH ₂ SEt) ⁺ /Ni(PPh ₂ CH ₂ CH ₂ SEt) ²	Ie	-0.65	- 1.14 ^{d,i}	-0.60	(-0.05)
Ni(Triphos)PPh ₃ ⁺ /Ni(Triphos)PPh ₃		-0.30	$-0.10^{c,h}$	-0.10	(-0.20)

 $^{a}\Delta E_{0} = E_{0}^{calc} - E_{0}^{SHE}$

^bSCE = the standard calomel electrode
^cSHE = the standard hydrogen electrode
^dAg/AgNO₃ = the standard silver electrode
^eFor the lowest energy conformers (Table S1)
^fRef. [72]
^gRef. [79]
^hRef. [28]
ⁱRef. [10]
^jRef. [85]

at the calculated reduction potentials of these complexes in Table 5, one can see that the reduction potentials of nickel complexes are in most cases much higher than for the base-on and base-off cobalamins and higher than for the methyl-cobalt dimethylglyoxime complexes with a pyridine ligand (baseon). This makes the radical reductive mechanism unlikely. On the other hand, the reduction potential of methyl cobalt glyoximate without pyridyne (base-off) is significantly higher than that for the base-on, contrary than in cobalamnis. Thus, the radical-reductive mechanism in principle could be possible for base-off glyoximate. This is probably not the case, because a pyridine or solvent molecule is attached to the cobalt atom in glyoximate complexes.

Conclusions

Several cobalt and nickel complexes involved in the methyl transfer reactions were examined with the DFT method using BP86 functional. The geometries, methyl binding energies and redox potentials of all the species were studied. For reduced cobalamins axial base undergo dissociation, which is consistent with experiment. In the base-off forms with water as an axial ligand, water molecule is linked by hydrogen bond to corrin nitrogen (CH₃CoCorH₂O). In methyl-free cobalamin (CoCorH₂O), the water molecule forms a hydrogen bond with cobalt atom.

Experimentally the five-coordinate structure for methylated nickel complex with PPh₂CH₂CH₂SEt ligand is suggested. Our calculations give small energy difference between five- and four-coordinate forms (1.6 kcal) which may imply fast interconversion between them.

There are noticeable differences in geometry, Co-CH₃ binding energies and redox potentials between cobalamin and dimethylglyoxime complexes, which indicates that chemical properties of these two systems are different. On the basis of the experimental redox potentials (-1.1 for CH₃Co(dmgBF₂)₂py/CH₃Co(dmgBF₂)₂py⁻ redox couple and -0.1 for Ni(Triphos)PPh₃⁺/Ni(Triphos)PPh₃), it was suggested that the reaction (2) takes place according to the S_N2 mechanism [28]. In the case of radical mechanism reduction of CH₃Co(dmgBF₂)₂py by Ni(Triphos)PPh₃ would be required. Our calculated redox potentials confirm such a statement, the calculated redox potential are equal to -0.99 V and -0.3 V, respectively.

Reaction (2) is fast [28] which can be attributed to the fact that the binding energy of methyl in the $CH_3Ni(Triphos)^+$ complex is about 10 kcal/mol higher than in the $CH_3Co(dmgBF_2)_2py$ complex.

Acknowledgments The Gaussian16 calculations were carried out in the Wroclaw Centre for Networking and Supercomputing, WCSS, Wroclaw, Poland, http://www.wcss.wroc.pl, under calculational Grant No. 18. Compliance with ethical standards This study complied with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Matthews RG, Koutmos M, Datta S (2008) Cobalamin-dependent and cobamide—dependent methyltransferases. Curr Opin Struct Biol 18:658–666. https://doi.org/10.1016/j.sbi.2008.11.005
- Matthews RG (2009) Cobalamin– and corrinoid– dependent enzymes. Met Ions Life Sci 6:53–114. https://doi.org/10.1039/BK 9781847559159-00053
- Jensen KP, Ryde U (2009) Cobalamins uncovered by modern electronic structure calculations. Coord Chem Rev 253:769–778. https://doi.org/10.1016/j.ccr.2008.04.015
- Svetlitchnaia T, Svetlitchnyi V, Meyer O, Dobbek H (2006) Structural insights into methyl transfer reactions of a corrinoid iron-sulfur protein involved in acetyl–CoA synthesis. Proc Natl Acad Sci U S A 103:14331–14336. https://doi.org/10.1073/pnas. 0601420103
- Ragsdale S, Pierce E (2008) Acetogenesis and the Wood-Ljungdahl pathway of CO₂ fixation. Biochim Biophys Acta 1784:1873–1898. https://doi.org/10.1016/j.bbapap.2008.08.012
- Ragsdale S (2004) Life with carbon monoxide. Crit Rev Biochem Mol Biol 39:165–195. https://doi.org/10.1080/10409230490496 577
- Drennan CL, Doukov TI, Ragsdale S (2004) The metalloclusters of carbon monoxide dehydrogenase/acetyl-CoA synthase: a story in pictures. J Biol Inorg Chem 9:511–515. https://doi.org/10.1007/s00775-004-0563-y
- Lindahl PA (2004) Acetyl-coenzyme A synthase: the case for a Ni_p(0) - based mechanism of catalysis. J Biol Inorg Chem 9:516–524. https://doi.org/10.1007/s00775-004-0564-x
- Gencic S, Grahame D (2008) Two separate one–electron steps in the reductive activation of the a cluster in subunit beta of the acds complex in methanosarcina thermophila. Biochemistry 47:5544–5555. https://doi.org/10.1021/bi7024035
- Hsiao YM, Chojnacki SS, Hinton P, Reibenspies JH, Darensbourg MY (1993) Organometallic chemistry of sulfur/phosphorus donor ligand complexes of nickel(II) and nickel(0). Organometallics 12:870–875. https://doi.org/10.1021/om00027a041
- Tan X, Surovtsev IV, Lindahl PA (2006) Kinetics of CO insertion and acetyl group transfer steps, and a model of the acetyl–CoA synthase catalytic mechanism. J Am Chem Soc 128:12331–12338. https://doi.org/10.1021/ja0627702
- Seravalli J, Kumar M, Ragsdale S (2002) Rapid kinetic studies of acetyl-CoA synthesis: evidence supporting the catalytic intermediacy of a paramagnetic NiFeC species in the autotrophic Wood-Ljungdahl pathway. Biochemistry 41(6):1807–1819
- Seravalli J, Ragsdale S (2008) Pulse–chase studies of the synthesis of acetyl—CoA by carbon monoxide dehydrogenase/acetyl–CoA

synthase: evidence for a random mechanism of methyl and carbonyl addition. J Biol Chem 283:8384–8394. https://doi.org/10. 1074/jbc.M709470200

- Ito M, Kotera M, Matsumoto T, Tatsumi K (2009) Dinuclear nickel complexes modeling the structure and function of the acetyl CoA synthase active site. Proc Natl Acad Sci U S A 106:11862–11866. https://doi.org/10.1073/pnas.0900433106
- Rigo P, Bressan M, Basato M (1979) Nickel(0) complexes with the hybrid bidentate ligand 1-(thioethyl)-2-(diphenylphosphino)ethane. Synthesis and catalytic properties of the related nickel hydride derivative. Inorg Chem 18:860–863. https://doi.org/10.1021/ic50193a064
- Stavropoulos P, Muetterties MC, Carrie M, Holm RH (1991) Structural and reaction chemistry of nickel complexes in relation to carbon monoxide dehydrogenase: a reaction system simulating acetyl-coenzyme A synthase activity. J Am Chem Soc 113:8485– 8492. https://doi.org/10.1021/ja00022a041
- Kitto HJ, Rae AD, Webster RD, Willis AC, Wild SB (2007) Synthesis, structure, and electrochemistry of di- and zerovalent nickel, palladium, and platinum monomers and dimers derived from an enantiopure (S, S)-tetra(tertiary phosphine). Inorg Chem 46:8059–8070. https://doi.org/10.1021/ic700912q
- Matsumoto T, Ito M, Kotera M, Tatsumi K A dinuclear nickel complex modeling of the nid(ii)-ni p(i) state of the active site of acetyl coa synthase, Dalton Transactions 39
- Ram MS, Riordan CG, Yap GPA, Liable-Sands L, Rheingold AL, Marchaj A, Norton JR (1997) Kinetics and mechanism of alkyl transfer from organocobalt(III) to nickel(I): implications for the synthesis of acetyl coenzyme A by CO dehydrogenase. J Am Chem Soc 119:1648–1655. https://doi.org/10.1021/ ja963061z
- Golden ML, Whaley CM, Rampersad MV, Reibenspies JH, Hancock RD, Darensbourg MY (2005) N₂S₂Ni metallodithiolate complexes as ligands: structural and aqueous solutionquantitative studies of the ability of metal ions to form M-S-Ni bridges to mercapto groups coordinated to nickel(II). Implications for acetyl coenzyme A synthase. Inorg Chem 44:875–883. https://doi.org/ 10.1021/ic0489701
- Webster CE, Darensbourg MY, Lindahl PA, Hall MB (2004) Structures and energetics of models for the active site of acetyl– coenzyme A synthase: role of distal and proximal metals in catalysis. J Am Chem Soc 126:3410–3411. https://doi.org/10.1021/ ja038083h
- Harrop TC, Olmstead MM, Mascharak PK (2006) Synthetic analogues of the active site of the A-cluster of acetyl coenzyme A synthase/CO dehydrogenase: syntheses, structures, and reactions with CO. Inorganic Cdrohemistry 45:3424–3436. https://doi.org/10.1021/ic0520465
- Dougherty WG, Rangan K, O'Hagan MJ, Yap GP, Riordan CG (2008) Binuclear complexes containing a methylnickel moiety: relevance to organonickel intermediates in acetyl–coenzyme A synthase catalysis. J Am Chem Soc 130:13510–13511. https://doi. org/10.1021/ja803795k
- Krishnan R, Riordan CG (2004) Cys-gly-cys tripeptide complexes of nickel: binuclear analogues for the catalytic site in acetyl coenzyme A synthase. J Am Chem Soc 126:4484–4485. https://doi.org/10.1021/ja038086u
- 25. Green KN, Brothers SM, Jenkins RM, Carson CE, Grapperhaus CA, Darensbourg MY (2007) An experimental and computational study of sulfur-modified nucleophilicity in a dianionic NiN₂S₂ complex. Inorg Chem 46:7536–7544. https://doi.org/10.1021/ic700878y
- Schenker R, Mock MT, Kieber-Emmons MT, Riordan CG, Brunold TC (2005) Spectroscopic and computational studies on [Ni(tmc)CH₃]OTf: implications for Ni-methyl bonding in the

A cluster of acetyl–CoA synthase. Inorg Chem 14:3605–3617. https://doi.org/10.1021/ic0483996

- 27. Schenker R, Kieber-Emmons MT, Riordan CG, Brunold TC (2005) Spectroscopic and computational studies on the trans-Mu-1,2-peroxo-bridged dinickel(II) species [Ni(tmc)₂(O₂)](OTf)₂: nature of end-on peroxo-nickel(II) bonding and comparison with peroxo-copper(II) bonding. Inorg Chem 44:1752–1762
- Eckert NA, Dougherty WG, Yap GPA, Riordan CG (2007) Methyl transfer from methylcobaloxime to (Triphos)Ni(PPh₃): relevance to the mechanism of acetyl–coenzyme A synthase. J Am Chem Soc 129:9286–9287. https://doi.org/10.1021/ja0720630
- Kozlowski PM, Kamachi T, Kumar M, Yoshizawa K (2012) Initial step of B₁₂-dependent enzymatic catalysis: energetic implications regarding involvement of the one-electron-reduced form of adenosylcobalamin cofactor. J Biol Inorg Chem 17:293– 300. https://doi.org/10.1007/s0077501108503
- 30. Zhou D-L, Walder P, Scheffold R, Walder L (1992) S_N2 or electron transfer? A new technique discriminates the mechanisms of oxidative addition of alkyl halides to corrinatoand porphyrinatocobalt(I). Helvetica Chimica Acta 75:995–1011. https://doi.org/10.1002/hlca.19920750403
- Banerjee S, Ragsdale S (2003) The many faces of vitamin B12: catalysis by cobalamin-dependent enzymes. Ann Rev Biochem 72:209–247. https://doi.org/10.1146/annurev.biochem.72.121801. 161828
- Jensen KP, Ryde U (2003) Theoretical prediction of the Co-C bond strength in cobalamins. J Phys Chem A 107:7539–7545. https://doi.org/10.1021/jp027566p
- 33. Dölker N, Maseras F, Lledös A (2001) A density functional study on the effect of the trans axial ligand of cobalamin on the homolytic cleavage of the Co—C bond. J Phys Chem B 105:7564–7571. https://doi.org/10.1021/jp010144f
- Andruniow T, Zgierski MZ, Kozlowski PM (2001) Theoretical determination of the Co-C bond energy dissociation in cobalamins. J Am Chem Soc 123:2679–2680. https://doi.org/10.1021/ ja0041728
- Jensen KP, Ryde U (2002) The axial N-base has minor influence on Co–C bond cleavage in cobalamins. J Mol Struct 585:239–255. https://doi.org/10.1016/S0166-1280(02)00049-0
- 36. Emelyanova N, Sanina AF, Shestakov DFT (2013) calculations of the redox potentials for the nitrosyl complexes [Fe₂(l-RS)₂(NO)₄] with R 5 alkyl. Int J Quantum Chem 113:740–744. https://doi.org/10.1002/qua.24063
- 37. Emelyanova N, Sanina N, Krivenko A, Manzhos R, Bozhenko K, Aldoshin S (2013) Comparison of pure and hybrid DFT functionals for geometry optimization and calculation of redox potentials for iron nitrosyl complexes with 1–SCN bridging ligands. Theor Chem Accounts 132:1316–1318. https://doi.org/10.1007/s002140 1213166
- Castro L, Bühl M (2014) Calculations of one-electron redox potentials of oxoiron(IV) porphyrin complexes. J Chem Theory Comput 10:243–251. https://doi.org/10.1021/ct400975w
- 39. Gaussian 16, Revision B.01, Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Scalmani G, Barone V, Petersson GA, Nakatsuji H, Li X, Caricato M, Marenich AV, Bloino J, Janesko BG, Gomperts R, Mennucci B, Hratchian HP, Ortiz JV, Izmaylov AF, Sonnenberg JL, Williams-Young D, Ding F, Lipparini F, Egidi F, Goings J, Peng B, Petrone A, Henderson T, Ranasinghe D, Zakrzewski VG, Gao J, Rega N, Zheng G, Liang W, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Throssell K, Montgomery JA Jr, Peralta JE, Ogliaro F, Bearpark MJ, Heyd JJ, Brothers EN, Kudin KN, Staroverov VN, Keith TA, Kobayashi R, Normand J, Raghavachari K, Rendell AP, Burant JC, Iyengar SS, Tomasi J, Cossi M, Millam JM, Klene M, Adamo C, Cammi

R, Ochterski JW, Martin RL, Morokuma K, Farkas O, Foresman JB, Fox DJ Gaussian, Inc., Wallingford CT, 2016

- Becke AD (1988) Density-functional exchange-energy approximation with correct asymptotic behavior. Phys Rev A 38:3098– 3100. https://doi.org/10.1103/PhysRevA.38.3098
- Perdew JP (1986) Density-functional approximation for the correlation energy of the inhomogeneous electron gas. Phys Rev B 33(12):8822–8824. https://doi.org/10.1103/PhysRevB.33. 8822
- Schäfer A, Huber C, Ahlrichs R (1994) Fully optimized contracted Gaussian basis sets of triple zeta valence quality for atoms Li to Kr. J Chem Phys 100:5829–5835. https://doi.org/10.1063/1.467 146
- Cossi M, Rega N, Scalmani G, Barone V (2003) Energies, structures, and electronic properties of molecules in solution with the C-PCM solvation model. J Comput Chem 24:669–681. https://doi.org/10.1002/jcc.10189
- Tomasi J, Mennucci B, Cammi R (2005) Quantum mechanical continuum solvation models. Chem Rev 105:2999–3073. https://doi.org/10.1021/cr9904009
- Himo F, Noodleman L, Blomberg MRA, Siegbahn PEM (2002) Relative acidities of ortho-substituted phenols, as models for modified tyrosines in proteins. J Phys Chem A 106:8757–8761. https://doi.org/10.1021/jp025646n
- Grimme S, Ehrlich S, Goerigk L (2011) Effect of the damping function in dispersion corrected density functional theory. J Comput Chem 32:1456–1465. https://doi.org/10.1002/jcc.21759
- 47. Grimme S, Antony J, Ehrlich S, Krieg H A consistent and accurate ab initio parametrization of density functional dispersion correction (DFT-D) for the 94 elements H-Pu. J. Chem. Phys. 132. https://doi.org/10.1063/1.3382344
- Stich TA, Brooks AJ, Buan NR, Brunold TC (2003) Spectroscopic and computational studies of Co³⁺-corrinoids: spectral and electronic properties of the B12 cofactors and biologically relevant precursors. J Am Chem Soc 125:5897–5914. https://doi.org/10.1021/ja029328d
- Jensen KP, Ryde U (2002) The axial N-Base is not important for Co-C bond cleavage in cobalamins. J Mol Struct (THEOCHEM) 585:239–255. https://doi.org/10.1016/S01661280(02)000490
- Dölker N, Morreale A, Maseras F (2005) Computational study on the difference between the Co-C bond dissociation energy in methylcobalamin and adenosylcobalamin. J Biol Inorg Chem 10:509–517. https://doi.org/10.1007/s00775-005-0662-4
- Hirao H (2011) Which DFT functional performs well in the calculation of methylcobalamin? Comparison of the B3LYP and BP86 functionals and evaluation of the impact of empirical dispersion correction. J Phys Chem A 115:9308–9313. https://doi.org/10.1021/jp2052807
- 52. Toscano PJ, Seligson AL, Curran MT, Scrobutt AT, Sonnerberger PC (1989) Cobalt-carbon bond disruption enthalpies: the first reliable measurement of a co-methyl bde via solution thermochemical methods. Inorg Chem 28:166. https://doi.org/10.1021/ic00300a038
- Dölker N, Maseras F, Lledós A (2001) A density functional study on the effect of the trans axial ligand of cobalamin on the homolytic cleavage of the Co-C bond. J Phys Chem B 105:7564– 7571. https://doi.org/10.1021/jp010144f
- Rutkowska Zbik D, Jaworska M, Witko M (2004) Application of the DFT theory to study cobalamin complexes. Struct Chem 15:431–435. https://doi.org/10.1023/B:STUC.0000037900. 67595.9f
- Kozlowski PM, Zgierski MZ (2004) Electronic and steric influence of trans axial base on the stereoelectronic properties of cobalamins. J Phys Chem B 108:14163–14170. https://doi.org/10.1021/ jp040373c

- Kumar N, Kuta J, Galezowski W, Kozlowski PM (2013) Electronic structure of on e-electron-oxidized form of the Methylcobalamin cofactor: Spin density distribution and Pseudo-Jahn-Teller effect. Inorg Chem 52:1762–1771. https://doi.org/10.1021/ic3013443
- 57. Kozlowski PM, Kumar M, Piecuch P, Li W, Bauman NP, Hansen JA, Lodowski P, Jaworska M (2012) The Cobalt-Methyl bond dissociation in Methylcobalamin: New benchmark analysis based on density functional theory and completely renormalized coupled-cluster calculations. J Chem Theory Comput 8:1870– 1894. https://doi.org/10.1021/ct300170y
- Hirao H, Kozlowski PM, Manoj K (2012) Co⁺. H interaction inspired alternate coordination geometries of biologically important cob(I)alamin: possible structural and mechanistic consequences for methyltransferases. J Biol Inorg Chem 17:1107–1121. https://doi.org/10.1007/s00775-012-0924-x
- Muckerman JT, Fujita E (2011) Theoretical studies of the mechanism of catalytic hydrogen production by a cobaloxime. Chem Commun 47:12456–12458. https://doi.org/10.1039/c1cc15330g
- Martin BD, Finke RG (1992) Methylcobalamin's full- vs. Half-Strength cobalt-carbon sigma bonds and bond dissociation enthalpies: Co-CH₃ homolysis rate Enhancement following oneantibonding-electron reduction of methlycobalamin. J Am Chem Soc 114:585–592. https://doi.org/10.1021/ja00028a027
- Kobylianskii IJ, Widner FJ, Kräutler K, Chen P (2013) Co–C bond energies in adenosylcobinamide and methylcobinamide in the gas phase and in silico. J Am Chem Soc 135:13648–13651. https://doi.org/10.1021/ja406676p
- Liptak MD, Brunold TC (2006) Spectroscopic and computational studies of Co¹⁺cobalamin: spectral and electronic properties of the superreduced B₁₂ cofactor. J Am Chem Soc 128:9144–9156. https://doi.org/10.1021/ja061433q
- Liptak MD, Datta S, Matthews RG, Brunold TC (2008) Spectroscopic study of the cobalamin-dependent methionine synthase in the activation conformation: effects of the Y1139 residue and S-adenosylmethionine on the B₁₂ cofactor. J Am Chem Soc 130:16374–16381. https://doi.org/10.1021/ja8038129
- 64. Kumar M, Kozlowski PM (2011) A biologically relevant $Co^{1+\cdots}H$ bond: possible implications in the protein-induced redox tuning of Co^{2+}/Co^{1+} reduction. Angew Chem 50:8702–8705. https://doi.org/10.1002/anie.201100469
- Andruniow T, Zgierski MZ, Kozlowski PM (2000) Density functional theory analysis of stereoelectronic properties of cobalamins. J Phys Chem B 104:10921–10927. https://doi.org/10.1021/jp000 810x
- 66. Galezowski W, Kuta J, Kozlowski PM (2008) DFT Study of Co-C bond cleavage in the neutral and one-electron-reduced alkyl-cobalt(III) phthalocyanines. J Phys Chem B 112:3177–3183. https://doi.org/10.1021/jp0769678
- Kuta J, Wuerges J, Randaccio L, Kozlowski PM (2009) Axial bonding in alkylcobalamins: DFT analysis of the inverse versus normal trans influence. J Phys Chem A 113:11604–11612. https://doi.org/10.1021/jp901397p
- Bühl M, Vinković Vrĉek I, Kabrede H (2007) Dehalogenation of chloroalkenes at cobalt centers. A model density functional study. Inorg Chem 26:1494–1504. https://doi.org/10.1021/om070027s
- Halpern J (1985) Mechanisms of coenzyme B₁₂-dependent rearrangements. Science 227:869–875
- Govender PP, Navizet I, Perry CB, Marques HM (2012) The cis influence of the corrin in vitamin B₁₂ models. Chem Phys Lett 550:150–155. https://doi.org/10.1016/j.cplett.2012.08.061
- Petrio M, Biarnés X, Kumar M, Rovira C, Kozlowski PM (2010) Reductive cleavage mechanism of Co-C bond in cobalamin dependent methionine synthase. J Phys Chem B 114:12965– 12971. https://doi.org/10.1021/jp1043738

- Birke RL, Huang Q, Spataru T, Gosser DKJ (2006) Electroreduction of a series of alkylcobalamins: mechanism of stepwise reductive cleavage of the Co-C bond. J Am Chem Soc 128:1922–1936. https://doi.org/10.1021/ja054479c
- Kepp K (2014) Co-C dissociation of adenosylcobalamin (Coenzyme B₁₂): Role of dispersion, induction effects, solvent polarity, and relativistic and thermal corrections. J Phys Chem A 118(34):7104–7117. https://doi.org/10.1021/jp503607k
- 74. Qu Z-W, Hansen A, Grimme S (2015) Co-C bond dissociation energies in cobalamin derivatives and dispersion effects: Anomaly or just challenging? J Chem Theory Comput 11:1037–1045. https://doi.org/10.1021/acs.jctc.5b00007
- 75. Lexa D, Saveant JM (1976) Electrochemistry of vitamin B_{12} . Role of the base-on/base-off reaction in the oxidoreduction mechanism of the B_{12r} - B_{12s} system. J Am Chem Soc 98:2652–2658. https://doi.org/10.1021/ja00425a039
- 76. Kozlowski PM, Kuta J, Galezowski W (2007) Reductive cleavage mechanism of methylcobalamin: elementary steps of Co-C bond breaking. J Phys Chem B 111:7638–7645. https://doi.org/10.1021/ jp066972w
- 77. Govender PP, Navizet I, Perry CB, Marques HM (2013) DFT Studies of trans and cis influences in the homolysis of the Co–C bond in models of the alkylcobalamins. J Phys Chem A 117:3057– 3068. https://doi.org/10.1021/jp311788t
- Han WG, Noodleman L (2011) DFT Calculations for intermediate and active states of the Diiron center with a tryptophan or tyrosine radical in Escherichia coli ribonucleotide reductase. Inorg Chem 50:2302–2320. https://doi.org/10.1021/ic1020127
- Lexa D, Saveant JM (1983) The electrochemistry of vitamin B₁₂. Acc Chem Res 16:235–243. https://doi.org/10.1021/ar00091a001

- De Tacconi NR, Lexa D, Saveant JM (1979) Electrochemistry of vitamin B-12. Kinetics and mechanisms in B_{12a} - B_{12r} oxido-reduction. J Am Chem Soc 101:467–472. https://doi.org/10. 1021/ja00496a034
- Lexa D, Saveant JM (1978) Electrochemistry of vitamin B₁₂.
 One-electron intermediates in the reduction of methylcobalamin and methylcobinamide. J Am Chem Soc 100:3220–3222. https://doi.org/10.1021/ja00478a048
- Fasching M, Schmidt W, Kräutler B, Stupperich E, Schmidt A, Kratky C (2000) Co(α—(1H—Imidazolyl)— Co-methylcob(III)amide: model for protein-bound corrinoid cofactors. Helvetica Chimica Acta 83:2295–2316. https://doi.org/ 10.1002/1522-2675(20000906)83
- Hung R, Grabowski J (1999) Listening to reactive intermediates: Application of photoacoustic calorimetry to vitamin B₁₂ compounds. J Am Chem Soc 121(6):1359–1364. https://doi.org/10.1021/ja9829620
- Luo L, Li G, Chen H, Fu S, Zhang S (1998) Laser-induced photoacoustic calorimetric determination of enthalpy and volume changes in photolysis of 5'-deoxyadenosylcobalamin and methylcobalamin. Dalton Trans, pp 2103–2108. https://doi.org/10.1039/ A708644J
- 85. Hu X, Brunschwig BS, Peters JC (2007) Electrocatalytic hydrogen evolution at low overpotentials by cobalt macrocyclic glyoxime and tetraimine complexes. J Am Chem Soc 129:8988–8998. https://doi.org/10.1021/ja067876b

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